

PCR DIRECT® Extracting Transport Medium

UFL105

PCR DIRECT Extracting Transport Medium

Instructions for Use

1. Product Description

PCR DIRECT Extracting Transport Medium (ETM) is intended for the inactivation, extraction, stabilization and transport of viral pathogens and host nucleic acids (DNA & RNA) from oral or nasal swabs (user-provided), or saliva. Extracted nucleic acids are suitable for either direct PCR/RT-qPCR or subsequent isolation using bead or column-based kits (user provided).

Arcis' PCR DIRECT reagents use a proprietary One-Step™ chemistry for rapid lysis, protection of nucleic acids and sequestration of potential inhibitors of PCR. These reagents allow ultra-rapid processing of swab or saliva samples, enabling users to go from prepared sample to downstream analysis in 1 pipette step, whilst maintaining high sensitivity and reproducibility in analytical results.

2. Materials Provided

Material Provided	Quantity	Number of Preps	Volume
PCR DIRECT ETM	1 Bottle	500*	124ml

×1



* 500 reactions based on standard saliva protocol volumes.

3. Storage Conditions

Store at ambient temperature. Do Not Freeze.

Bottles, vials and tubes should be capped when not in use.

4. Samples

PCR DIRECT ETM is optimized for preparation of PCR ready DNA or RT-qPCR ready RNA from nasal, oral swabs or saliva. User-provided swabs should be sterile and deemed appropriate for the application. Swabs, immediately after collection, should be placed into sterile collection tubes containing aliquoted PCR DIRECT ETM.

Saliva samples should be collected into dry tubes or receptacles. For saliva samples, donors should be instructed not to consume food or drink, or use gum, toothpaste for at least 30 minutes prior to sample collection. Saliva should not be mixed with other transport media or lysis reagents prior to the addition of the PCR DIRECT ETM as this may affect results.

5. Applications

Nucleic acids prepared with PCR DIRECT ETM have been successfully used as samples in molecular biology techniques including PCR, and RT-qPCR. For maximum sensitivity in downstream PCR, the reaction may be prepared with up to 50% sample by volume (e.g., up to 12.5µl sample in a 25µl reaction). Nucleic acid yield and processing efficiency may vary depending on the virus extracted. Samples prepared with the PCR DIRECT ETM should be used in the intended downstream processes within 4 days of preparation or frozen if intended for use after 4 days. Users should avoid repeated freeze thaw cycles.

Instructions for Use continued

6. Standard Swab Protocol

If the collected samples are for direct PCR analysis, we recommend preparing sample collection tubes ahead with 300-500µl of PCR DIRECT ETM per tube. If a bead- or column-based isolation kit will be used, 200-400µl of sample in PCR DIRECT ETM may be used for subsequent processing

1. Collect oral or nasal swabs according to laboratory procedure
2. Place swab head into PCR DIRECT ETM; swirl and rotate swab head against the tube wall at least 10 times, to extract the maximum amount of liquid from the swab head
3. Begin to remove swab, press swab head against the side of the collection tube
4. Shake tube vigorously at least ten times
5. Sample is ready for transport and subsequent downstream molecular analysis or processing

Note: the head of the swab should be fully submerged in a minimal volume of PCR DIRECT ETM. Complete immersion will ensure that the entire collected sample has been lysed. Minimizing the volume of PCR DIRECT ETM ensures the highest concentration of nucleic acids and supporting direct PCR testing methods.

7. Standard Saliva Protocol

Collect saliva sample according to laboratory procedure, in a dry tube without diluents/transport medium. This protocol may be adjusted for larger sample volumes by scaling the volume of PCR DIRECT ETM in proportion to the increase in sample volume.

1. Add 100µl of saliva to a 1.5ml low binding micro tube
2. Add 235µl of PCR DIRECT ETM to the sample with the same pipette, mix ten (10) times
3. Sample is ready for transport and subsequent downstream molecular analysis or processing

Note: Inadequate mixing in step 2 will result in lower sensitivity. Mixing by pipetting is preferred



This product is intended for laboratory research use only. Not for diagnostic use.



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